# **Acute Toxicity**

Test Substance Heavy Aromatic Distillate, CAS# 64742-48-9

None

Olefins Panel HPV Stream Name: Hydrotreated C8-C10

No guideline specified, comparable to standard study

Method

Method/guideline followed

Type (test type)

Acute LD50 **GLP** Yes 1984 Year

Species/Strain Rat. Fischer 344 Male and female 5/dose/group (4 groups)

No. of animals per sex per

dose

Vehicle

Route of administration

Oral gavage

**Test Conditions** Rats were dosed once with undiluted heavy aromatic distillate at 4.5, 5.0, 5.5 and

and clinical signs. Body weight was obtained at initiation, and after 7 and 14 days

Results

LD<sub>50</sub> with confidence limits.

Remarks

**Conclusions** 

(study author)

Data Quality Reliability

References

Other

Last changed

6.0 g/kg, and were observed for 14 days post dosing for mortality, moribundity

post dosing. Gross necropsies were performed on all rats at study termination.

There were no deaths attributed to test article administration, and therefore, the LD50 was not reached in either sex at the highest dose. Body weight was not

significantly changed over the 14-day observation period. Over the first week of study, there were instances of perianal soiling, dry material around the mouth, and soft feces. In sacrificed rats, there were no findings that could be attributable to

test material administration.

The LD50 was not reached at the highest dose of 6.0 g/kg.

1. Reliable without restrictions

Rausina, G. 1984. Acute oral toxicity study in albino rats using heavy aromatic

distillate. Proj. # 2049. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil

Chemicals Co., Houston, TX.

Rev 6/25/2001 (Prepared by a contractor to the Olefins Panel)

# **Acute Toxicity**

#### Test Substance

Heavy aromatic distillate, CAS# 64742-48-9. No composition or purity analysis reported, refer to sponsor. Olefins Panel HPV Stream Name: Hydrotreated C8-

C10

#### Method

Method/guideline followed

Type (test type)

GLP Year

Species/Strain

Sex

No. of animals per sex/dose

Vehicle

Route of administration

No guideline specified but comparable to standard study

Acute LC50

Yes 1983

Rat, Fischer 344 Male and female

5

filtered air

whole body inhalation

**Test Conditions** 

Six groups of 10 rats (5M, 5F/group, 12-16 wks old, 145-307g) were individually housed and exposed in stainless steel/glass inhalation chambers to aerosolized test article or filtered air for 4 hours, followed by 14 days of observation post exposure for clinical signs, morbidity and death. Non-fasted rats were sacrificed on day 14 and necropsied for gross lesions. Nominal chamber concentrations (g/m³) were 0.0, 12.2, 24.8, 25.8, 19.6, and 99 (uncorrected for large particle condensation), but actual chamber concentrations were 0, 6.0, 7.6, 8.6, 9.1, and 11.2 as determined by gas chromatography. Probit analysis was used to estimate an LC50.

#### Results

LC<sub>50</sub> with confidence limits. Remarks

LC50 was estimated to be 8.5 g/m<sup>3</sup> (actual chamber concentration). There was a large difference between nominal and actual concentrations in the inhalation chambers that was not addressed in the report. In the analyses, exposure concentration was estimated by comparing peak height (rather than peak area) with that of the neat sample. The method of calculating chamber concentration of test article in ppm was not reported (Comment by contractor). All animals in the high dose group died during exposure with congestion of lungs and nasal turbinates with red discharge. Six animals died in groups 4 and 5 during exposure, and were found with gas in the G.I. tract. Mean body wt of males and females decreased by day 7 but then increased over the remaining 7 days. Most rats exhibited nasal and ocular discharges, and in the higher dose groups showed signs of CNS effects, (hyperexcitability, twitching, circling) that . were absent by day 2. Other clinical effects were absent by day 7. No test article related gross pathological lesions were observed.

**Conclusions** 

LC50 was  $8.5 \text{ g/m}^3$ .

<u>Data Quality</u> Reliability

1. Reliable without restrictions. This study is acceptable for range-finding since the concentrations employed yielded a dose response curve covering the full range of biological response (0-100% fatalities).

<u>References</u>

Goode, J.W. 1983. LC50 Inhalation toxicity study in rats using heavy aromatic distillate. Proj. # 2050. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co., Houston, TX

Other

Last changed

Rev. 7/2/2001 (Prepared by a contractor to the Olefins Panel)

In vitro mammalian cell forward mutation

Chinese hamster ovary (CHO) cell culture

# **Genetic Toxicity - in Vitro**

Test Substance

Test substance

Heavy Aromatic Distillate, Gulf. CAS #64742-48-9. Water-white liquid with characteristic aromatic odor. Composition analysis, purity and stability referred to sponsor.

Olefins Panel HPV Stream Name: Hydrotreated C8-C10

Standard method based on Hsie et al. (1981), O'Neill & Hsie (1979)

Rat liver (S9) fraction purchased from Litton Bionetics, Kensington, MD

1.0mg S9 fraction/ml treatment medium (0.3ml S9 fraction in 3 ml medium/flask)

**Method** 

Method/guideline followed

Type

System of testing

GPYear

Species/Strain

CHO-K-1 heterozygous for hypoxanthine-guanine phosphoribosyl transferase (HGPRT+/-) from Oak Ridge National Laboratory, TN.

Yes 1984

Yes

Metabolic activation

Species and cell type

Quantity

Induced or not induced

Aroclor 1254 induced (treatment not specified) Cytotoxicity, final conc. (trial 2): 128, 256, 512,  $1024ug/ml \pm S9$ : Concentrations tested

> Mutagenicity, final conc. (trial 2): 64, 128, 256, 512, 750, 1024µg/ml –S9; 128, 256, 512, 1024, 1500, 2048µg/ml +S9, all diluted in 10% Pluronic polyol F68 (prepared in dionized

water, mol. wt. 8350)

Statistical Methods

Frequency of mutant colonies per million clonable cells was calculated and comparisons of treated cultures with vehicle controls made on transformed data using a two-tailed t-test (Irr & Snee, 1979). Criteria for positive results were significant (p<0.05) increase in mutant colonies (HGPRT+/- HGPRT-/-) at any dose level and a dose related response. If only

one criterion is met, results are considered equivocal.

Remarks for Test Conditions

Sufficient Heavy Aromatic Distillate (HAD) was weighed separately for each dose level into 10 ml volumetric flasks, 1.8ml of 10% F68 added per ml of final volume and medium (Ham's F-12 without hypoxanthine) added as required to achieve final 10ml volume for testing. All dosing preparations were vortexed just after addition of medium and just prior to use when 20µl of each preparation was added to 3ml treatment medium/culture vessel. All cultures were incubated at 37°C in 5% CO2 enriched, humidified atmosphere. Positive control mutagens were ethyl methanesulfonate (100µg/ml) for -S9 cultures, and benzo(a)pyrene (4μg/ml) for +S9 cultures. For cytotoxicity, each dose group was composed of 2 flasks, one -S9, one +S9, negative controls  $\pm S9$ , seeded with  $5 \times 10^{3}$  cells on day 1. Cultures were exposed to test compound for 5 hours on day 2. On day 3, cells were trypsinized and counted with a Coulter Model ZB, then 200 cells were transferred into each of 3 60mm culture dishes. These viability plates were incubated until day 10, fixed in methanol and stained with Giemsa. Colonies were counted visually or with an Artek Model 981 colony counter. Absolute survival = total colony count ÷ number of cells seeded/flask. Relative survival = absolute survival in treated cultures ÷ vehicle control survival. Acceptable survival level is at least 10%. For mutagenicity, cells were seeded on day 1 into 6 flasks/dose group, 3-S9, 3+S9; on day 2 approximately 10<sup>6</sup> cells were exposed to HAD for 5 hours. Vehicle control had 12 flasks, 6-S9, 6+S9. On day 3, cultures with excessive cytotoxicity were discarded. From remaining cultures, 200 cells were seeded to each of 4 viability plates/dose level; incubated to day 10, fixed with methanol, stained with Giemsa, and colonies counted for survival. Expression cultures (10<sup>5</sup>-10<sup>6</sup> cells/one dish/dose) were seeded on day 3; subcultured three times until day 10 when 200 cells were seeded on each of 4 viability plates/dose and 2x10<sup>5</sup> cells seeded on each of 5 mutagenicity plates/dose with selective medium containing 10<sup>-5</sup>M 6-thioguanine to allow expression of HGPRT mutation. Cultures were incubated undisturbed until day 17 when they were fixed and stained. For mutagenicity, a ratio of total colony counts in mutagenicity plates over absolute survival in viability plates was calculated for each treatment group. Frequency of

mutant colonies/million clonable cells was calculated and statistical comparisons with negative control data were made.

#### Results

Genotoxic effects

First trial was aborted after post treatment cell counts in the mutagenicity assay, because of disparity in cytotoxic response between cytotoxicity aspect and mutagenicity aspect of the trial. No cytotoxicity was seen in the mutagenicity aspect. Results of the second cytotoxicity trial demonstrated cytotoxicity at 512  $\mu$ g/ml and higher in the –S9 portion and only a slight decrease in cell count at 1024  $\mu$ g/ml in the activated portion but no cytotoxicity in colony count after treatment. The second mutagenicity trial indicated post treatment cell death in –S9 cultures beginning at 64  $\mu$ g/ml with significant effects at 256  $\mu$ g/ml and 100% toxicity at 750  $\mu$ g/ml. In +S9 cultures, slight cell toxicity was seen at 1500  $\mu$ g/ml and above. For both activated and non-activated cultures, no significant dose-related responses or significant increase in mutant frequency were observed at any dose level of HAD. Positive control compounds responded appropriately (EMS 5.2 fold increase and B(a)P 2.8 fold increase over vehicle control).

# **Conclusions**

(contractor)

Mutagenicity was not observed at any HAD dose level tested with or without metabolic activation. Heavy Aromatic Distillate does not induce gene point mutations in the CHO/HGPRT test under conditions of this assay.

# Data Quality

Reliabilities

1. Reliable without restrictions. Study conforms to standard design. GLPs have been followed

#### **Reference**

Papciak, M.S., Goode, J.W. 1984. CHO/HGPRT test: Heavy Aromatic Distillate. Proj. #2054. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co., Houston, TX

Hsie, A.W. et al. 1981. Mut. Res. 86: 193-214

O'Neill, J.P. and Hsie, A.W. 1979. Banbury Report 2: 55-63 Irr, J.D. and Snee, R.D. 1979. Banbury Report 2: 263-275.

#### **Other**

Last changed

Rev. 6/25/2001 (Prepared by a contractor to the Olefins Panel)

#### **Genetic Toxicity - in Vitro**

Test Substance

Test substance

Heavy Aromatic Distillate, Gulf. CAS #64742-48-9. Water-white liquid with characteristic aromatic odor. Composition analysis, purity and stability referred to sponsor.

Olefins Panel HPV Stream Name: Hydrotreated C8-C10

**Method** 

Method/guideline followed

Type

System of testing

GLP Yes Year 1984

Species/Strain BALB/3T3-A31-1-1 from T. Kakunaga, National Cancer Inst., 1982

Metabolic activation No
Species and cell type NA
Quantity NA
Induced or not induced NA

Concentrations tested Cytotoxicity: 8, 16, 32, 64, 128, 256, 512, 1024, 2048, 5000µg/ml;

In vitro cell transformation

Mouse embryo cells

Transformation: 16, 32, 64, 200µg/ml, all diluted in 10% Pluronic <sup>®</sup> polyol F68 (prepared in

Standard method based on Cortesi et al (1983), Dunkel et al (1981), Reznikoff et al (1973)

deionized water, mol. wt. 8350, 80% hydrophilic).

Exposure period 2 days Statistical Methods None e

None employed. Criteria for positive response were a two-fold increase in type III foci at the highest dose over vehicle control (at least 2 type III foci if vehicle control had none) with or without a dose related response, or a two-fold increase at two or more consecutive doses. Test is equivocal if two-fold increase occurred at any one level other than the

highest acceptable dose.

Remarks for Test Conditions

Sufficient Heavy Aromatic Distillate (HAD) was weighed separately for each dose level, 0.45ml of 10% F68 added per ml of final volume and medium (Eagles MEM with 10% heat-inactivated fetal calf serum) added as required to achieve final volume for testing. Test preparations were mixed just prior to addition to cultures at 50µl to each 5 ml culture. All cultures were incubated at  $37^{0}$ C in 5% CO2 enriched humidified atmosphere. For cytotoxicity, 2 plate cultures/dose group, 2 plate cultures for vehicle F68 or medium negative control were seeded with  $1\times10^{4}$  cells/plate in day 1, exposed on days 2-3, trypsinized and counted with a Coulter Model ZB on day 4 for at least 20% survival. For transformation, 15 flasks  $(1\times10^{4}$  cells/flask/dose group)) and two cloning flasks (100 cells per flask/dose group) were seeded on day 1, exposed on days 2-3 and culture medium changed on day 4. For transformation flask cultures, medium continued to be changed weekly to day 29. Positive control was 3-methylcholanthrene (1µg/ml). Cloning flask cultures were fixed, stained, and counted visually on day 8 to determine cloning efficiency (avg. number colonies/plate  $\div$  100 cells seeded). Flask cultures were fixed and stained on day 29 for focus counting and evaluation. Transformation frequency = total type III foci  $\div$ 

total flasks/dose group.

Results

Genotoxic effects

In the first trial, HAD induced toxicity in BALB/3T3 cells after two days exposure beginning at  $32\mu g/ml$  (59.9% relative survival), increasing with dose level to 2.9% relative survival at  $5000\mu g/ml$ . The first trial was discarded due to loss of many cultures (27/105) due to contamination. Results of the second transformation trial indicated no treatment related cell transformation induced by HAD. Toxicity was evident  $32\mu g/ml$  (67.2% relative cloning efficiency), increasing sharply at  $200\mu g/ml$  (28.8% cloning efficiency). Positive and negative controls gave expected results.

Conclusions (contractor)

Heavy Aromatic Distillate did not induce transformation in BALB/3T3 cells at any dose level under conditions of this assay.

<u>Data Quality</u> Reliabilities

1. Reliable without restriction. Study conforms to standard design. GLPs have been followed.

<u>Reference</u>	Brecher, S., Good, J.W. 1984. BALB/3T3 transformation test: Heavy Aromatic Distillate. Proj. #2057. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co, Houston, TX Cortesi, E. et al. 1983. Teratogenesis, Carcinogenesis, Mutagenesis 3: 101-110. Dunkel, V.A. et al. 1981. J. Nat'l Cancer Inst. 67: 1303-1315. Reznikoff, C.A. et al. 1973. Cancer Res. 3239-3249.
<u>Other</u>	
Last changed	4/11/2001 (prepared by a contractor to the Olefins Panel).

# **Genetic Toxicity - in Vitro**

Test Substance

Test substance

Heavy Aromatic Distillate, Gulf. CAS #64742-48-9. Water white liquid with

characteristic aromatic odor. Composition analysis, purity and stability referred to sponsor.

Olefins Panel HPV Stream Name: Hydrotreated C8-C10

**Method** 

Method/guideline followed

Type

System of testing

GPYear

Species/Strain

Metabolic activation Species and cell type

Quantity

Induced or not induced

Concentrations tested

Exposure period Statistical Methods

Remarks for Test Conditions

Results

Genotoxic effects

**Conclusions** (contractor)

Standard method based on Williams et al. (1977,1982)

In vitro mammalian cell DNA repair assay

Unscheduled DNA Synthesis (UDS) in primary hepatocyte cultures.

Yes 1984

Fischer 344 male rat (13-14 wks old) – 1 rat per test

NA NA NA

Range-finding: 8, 16, 32, 64, 128, 256, 512, 1024, 2048, 5000 µg/ml: UDS assay 10, 40,

100, 200 µg/ml; all diluted in 10% Pluronic polvol F68 (prepared in deionized water, mol.

wt 8350, 80% hydrophilic)

18 hours

None employed. Criteria for positive response are incorporation of radioactive precursor (<sup>3</sup>H-thymidine) in cells that are not normally synthesizing DNA, indicating repair of damage. A positive response is defined as a mean net nuclear grain count at any treatment level that exceeds concurrent negative control by at least 6 grains/nucleus; negative control

value must not exceed 5 grains. A positive response need not be dose related.

Sufficient Heavy Aromatic Distillate (HAD) was weighed separately for each dose level. 0.45ml of 10% F68 added per ml of final volume and sufficient medium (Williams Medium E with 10% fetal bovine serum and insulin) added to achieve final volume. Test preparations were mixed just prior to addition at 30µl to each 3 ml culture. The conc. of <sup>3</sup>H-thymidine (1/4) ife 12.5 yrs.) used in these assays was 1mCi/ml. All cultures were incubated at 37°C in 5% CO<sub>2</sub> enriched humidified atmosphere. For range-finding, primary hepatocytes derived from freshly perfused rat liver were seeded (approx. 1x10<sup>5</sup> cells/ml) into treatment vessels, exposed to test material for 18 hours (2 cultures/dose level; 2 untreated cultures, and two vehicle (F68) control cultures), then fixed in formalin and stained with trypan blue for viability determination. At least 50% viability needed for the assay. In the UDS assay, 1x10<sup>5</sup> cells/ml were seeded into coverslip cultures, exposed to <sup>3</sup>H-thymidine and test substance for 18 hours (3 cultures/dose level). Positive control was 2-acetyl aminofluorene (0.2µg/ml). Cells growing on coverslips were rinsed, fixed and glued to microscope slides on day 2. On day 3, slides were dipped in autoradiographic emulsion and stored in the dark at 2-8°C. Autoradiographs were developed, stained and coverslipped on day 14. Numbers of grains overlying 50 randomly selected nuclei/slide were counted. The highest of 3 cytoplasmic grain counts/cell were subtracted and this number was divided by a conversion factor of 2, to obtain net nuclear grain count. Avg. net nuclear grain count/slide (sum of net nuclear grain count ÷ 50) and mean net nuclear

HAD induced toxicity in primary hepatocytes beginning at 32-64µg/ml (72-80% relative viability) after 18 hours exposure, which increased with dose levels to 2% viability at 5000µg/ml. HAD did not cause unscheduled DNA synthesis at any dose level. Positive and negative controls gave expected results.

grain count (avg. net nuclear grain count/slide ÷3) were calculated.

Unscheduled DNA synthesis was not observed in primary culture of rat hepatocytes at any dose level of Heavy Aromatic Distillate, indicating that this material does not damage DNA under conditions of this assay.

Data Quality Reliabilities	1. Reliable without restrictions. Study conforms to standard design. GLPs have been followed.
<u>Reference</u>	Brecher, S., Goode, J.W. 1984. Hepatocyte primary culture/DNA repair test of heavy aromatic distillate. Proj. #2056. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co., Houston, TX Williams, G.M. 1977. Cancer Res. 37: 1845-1851 Williams et al. 1977. In Vitro 13: 809-817 Williams et al. 1982. Mut. Res. 97:359-370
<u>Other</u>	
Last changed	4/11/2001 (Prepared by a contractor to the Olefins Panel)

# **Genetic Toxicity - in Vivo**

Test Substance

Remarks

Heavy Aromatic Distillate, Gulf CAS #64742-48-9. Water white liquid with aromatic

odor. Compositional analysis, purity and stability referred to sponsor.

Olefins Panel HPV Stream Name: Hydrotreated C8-C10

<u>Method</u>

Method/guideline followed

Type GLP

Year Species

Strain/Sex

Route of administration Doses/concentration levels Exposure period

Statistical methods

Remarks for Test Conditions.

Results

Genotoxic effects NOAEL (NOEL) LOAEL (LOEL)

<u>Conclusions</u> (study authors)

<u>Data Quality</u> Reliabilities

References

Comparable to standard assay

Mammalian bone marrow erythrocyte micronucleus

Yes 1984 Mouse

Crl:CD<sup>®</sup>-1 (ICR) BR Swiss: Male and female: Range finding (RF): 2M, 2F/group;

Micronucleus: 10M, 10F/group; 15M, 15 F in 1 group

Oral gavage

RF: 0, 1.25, 2.5, 5.0 g/kg in corn oil: Micronucleus: 0, 0.625, 1.25, 2.5 g/kg in corn oil

1 dose/day for 2 days; 1 group at 2.5 g/kg 1 dose, 1 day only

Values from treated groups for daily mean body weights, group means and std. dev. for polychromatic erythrocytes (PCEs) with micronuclei (MN), and group mean ratios of PCE to normochromatic erythrocytes (NORMs) were calculated and compared with vehicle control values by Student's t-test. Positive response was indicated by statistically significant (p<0.05) increases in micronucleated PCE at any dose level with a dose related response evident. Results were considered equivocal if only one of these criteria was met.

Heavy Aromatic Distillate (HAD) dosing solutions were prepared fresh for each day of dosing –12.5 g HAD (RF) or 6.25 g HAD (micronucleus) mixed with corn oil to make 50 ml, blended by shaking. Based on results of the range finding study, three groups of mice were given HAD by oral gavage daily for two days. All mice were weighed on day 1 and on day of sacrifice. One half of each treated group and vehicle control (5M, 5F) was killed on day 3 and the remainder on day 4. One group (15M, 15F), given 2.5 g/kg by gavage in a single dose for 1 day only, was killed on days 2, 3, 4 (5/sex/day). Positive control mice given cyclophosphamide (75 mg/kg) ip daily for 2 days were killed on day 3. Slides of femoral bone marrow smears were prepared, stained with May-Grunewald/Giemsa stain and examined microscopically. For each mouse, 1000 PCE and all associated mature erythrocytes (NORMs) were counted. Data collected included group mean body weights for each day, total PCEs, total NORMs, PCEs with MN, and NORMs with MN.

In range finding test, 1/2males and ½ females died at 5.0 g/kg dose level by day 3. In the micronucleus test, 1/10 females in the 2.5 g/kg dose group (2 days of dosing) died by day 4. All other mice survived to study sacrifice. Body wts were comparable to negative controls for both sexes in all treatment groups and positive controls. Treatment with HAD did not show any significant changes in micronucleus formation or in the ratio of PCE/NORM at any dose level. Average PCE/NORM ratio was 0.9% for all HAD treatment groups and negative control; ratio for positive control was 0.5%. NOEL (systemic) = 1.25 g/kg: NOEL (genetic) = 2.5 g/kg

Oral treatment of mice with Heavy AaromaticDistillate for 1 or 2 days at doses up to 2.5 g/kg did not cause increased frequency of micronucleated polychromatic erythrocytes in bone marrow of treated mice. Under these test conditions, Heavy Aromatic Distillate does not induce cytogenetic damage.

1. Reliable without restrictions. Study conforms to standard design. GLP followed.

Khan, S.H. and Goode, J.W. 1984. Micronucleus test in mouse bone marrow: Heavy

	Aromatic Distillate administered orally for 2 days. Proj. #2005. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co., Houston, TX
Other Last changed	Center, Franciscon, Francisco,
	Rev. 6/25/2001 (Prepared by a consultant to the Olefins Panel)

#### **Repeated Dose Toxicity**

Test Substance

Remarks

Heavy Aromatic Distillate, CAS # 64742-48-9. No analysis of purity or composition reported; referred to sponsor. Olefins Panel HPV Stream Name: Hydrotreated C8-C10

Method

Method/guideline followed

No guidelines specified, comparable to standard study Test type Subacute GLP Yes Year 1983 Species Rat

Strain Fischer 344

Route of administration Whole body inhalation

Duration of test 5 days

Doses/concentration levels  $0, 1.2, 2.7, 5.0 \text{ g/m}^3$ 

Males and females 5/sex/group Sex Exposure period 5 days

Frequency of treatment 6 hours/day

filtered air at 6 hrs/day for 5 days Control group and treatment

None

Post exposure observation period

Statistical methods

Analysis of Variance, Dunnett's test

**Test Conditions** 

Animals (13 weeks old at study initiation, 156-279g) were housed individually in screenbottom cages with automatic watering in rooms maintained at approx. 74°F with relative humidity of 50%, and 12 hour light/ dark cycle. Chow diet and water were provided ad lib except during exposure. Chamber concentrations were monitored by GC; peak areas were compared with those of neat test article standards. Rats were monitored twice daily for morbidity and mortality, and observed once daily for clinical signs. Body weights were measured at initiation and termination. Necropsies were performed for gross lesions.

Results

NOAEL (NOEL) LOAEL (LOEL)

Remarks

NOEL not determined

LOEL = 1.2 g/m<sup>3</sup> based on clinical observations: perianal staining, red material around nose/mouth, ocular porphyrin. (assessed by Reviewer). One female rat in the high dose group died during the initial exposure; all other rats survived until termination. Males and females exposed to 5.0g/m3, showed a dose related weight loss of approx. 8% after 5 days of dosing. Incidence of dry red material around nose/mouth, ocular porphyrin, clear discharge from the eyes, partially closed eyes and perianal staining occurred in all groups receiving test article. The two high dose groups showed purulent discharge from the eyes and bloody tears.

Reviewer comment: The total incidence of clinical observations increased in a manner related to exposure concentration.

**Conclusions** 

One death occurred during exposure of the high dose group. All other animals exhibited clinical signs that included ocular discharge, eye closure, and dry red material around the nose/mouth. Gross pathological lesions were not observed which could be directly attributable to test article administration.

Quality

Reliabilities

1. Reliable without restrictions.

References

Gordon, T. 1983. One week repeat dose inhalation toxicity study in the rat using heavy aromatic distillate. Proj. # 2062. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co., Houston, TX.

Other

Last changed

Rev. 7/3//2001 (Prepared by a contractor to the Olefins Panel)

# **Repeated Dose Toxicity**

Test Substance

Heavy Aromatic Distillate, CAS #64742-48-9. No composition or purity analysis reported;

refer to sponsor. Olefins Panel HPV Stream Name: Hydrotreated C8-C10

Remarks Method

Method/guideline followed No guideline specified; comparable to standard study.

Test type Subacute
GLP Yes
Year 1985
Species Rat
Strain Fischer 344

Route of administration Dermal
Duration of test 4 weeks

Doses/concentration levels 0.0, 0.5, 1.0, 1.5 g/kg in paraffin oil vehicle

Sex Males and female (10/sex/group), 72 days old at study initiation

Exposure period 6 hours/day

Frequency of treatment once/day, 5 days/week

Control group and treatment Paraffin oil, 2.18 ml/kg/day for 5 days/week

Post exposure observation period | No

Statistical methods Bartlett's test for homogeneity, Dunnett's test for homogeneous data; modified t-test for

non-homo geneous data.

Test Conditions

Animals were housed individually in suspended stainless steel cages with wire mesh bottoms and fronts equipped with an automatic watering system, in a room maintained at 76.1°F with relative humidity of 56.6% and 12 hour light/dark cycle. Chow diet was provided ad lib. Test article dilutions in paraffin oil (75% v/v) were prepared weekly. Doses of test article were administered over 10% of body surface to the backs of rats clipped free of hair and fitted with Elizabethan collars to reduce ingestion. After 6 hours, collars were removed and residual oil wiped off. Observations for mortality and moribundity were made twice/day, and for clinical signs at least once daily (on dosing days). Dermal responses were scored at initiation and then weekly. Body weight was measured at initiation and then weekly. Food consumption was determined weekly. At sacrifice, gross necropsy was performed, organs/tissues (19/rat) weighed and preserved. Slides prepared for histopathologic examination for the following tissues/organs of control and high dose groups: brain, spinal cord, heart, lungs, thymus, left kidney, right kidney, liver, spleen, sternum, lymph nodes, testes, skin, adrenal glands, urinary bladder, and peripheral nerve.

Results

NOAEL (NOEL)

LOAEL (LOEL)

NOEL not determined.

LOEL males = 0.5g/kg

Remarks

LOEL males = 0.5g/kg (increased total WBC count, assigned by reviewer) LOEL females = 1.5 g/kg (hematologic alterations, skin irritation, assigned by reviewer) No deaths or moribund rats were observed and no statistically or biologically significant differences in group mean body wt were noted at study termination. No clinical effects were observed that could be attributed to test article administration. Food consumption was significantly decreased in male rats given 1.5g/kg during wks 2 and 3, and in female rats given 1.0g/kg during wk 2. Severe erythema was observed in 1.5g/kg males and females by wk 3 which persisted for the duration of the study. At termination, moderate eschar formation was seen in 10/10 males and 7/10 females. Statistically significant changes in hematology and clinical chemistry parameters after 4 wks of dosing were: dose responsive increase in WBC (57-70%) of males and females in 1.5g/kg group; slight reduction of RBC in males and reduction of HGB and HCT of males and females given 1.5g/kg; elevated platelet counts (10-20%) in 1.5g/kg males and females, reduced total serum protein (10-13%) in 1.5g/kg males and females; reduced serum albumin (9-25%) in high dose males and females; dose responsive reduction in BUN (9-25%) in high dose animals. There were marked increases in segmented neutrophils (200-400%) and lymphocytes (20-30%) in males and females given 1.5g/kg, and an erratic but marked increase in atypical lymphocytes of males in low-high dose groups and increased eosinophils (485%) in 1.5g/kg males. There were several statistically significant but inconsistent changes in organ wt when expressed as

	absolute wt, or per 100 g body wt. but these were not perceived as being biologically significant. There were no histopathological effects noted except those in the skin.
Conclusions	Repeated application of heavy aromatic distillate to male and female rats caused severe skin irritation and significantly decreased food consumption and body wt. Gross and microscopic lesions produced at the site of application, included ulceration, acanthosis and hyperkeratosis. In both male and female rats, treatment with heavy aromatic distillate was associated with significantly elevated WBC counts and mild anemia associated with decreased RBC counts, hematocrit, level of hemoglobin in peripheral blood, and elevated platelet count. The elevated WBC count was related to elevated levels of neutrophils and lymphocytes.
<i>Quality</i> Reliabilities	2. Reliable with restrictions. No analysis of test material preparations in paraffin oil.
References	Zellers, J.E. 1985. Four week repeated dose dermal toxicity study in rats using heavy aromatic distillate. Proj. #2063. Gulf Life Sciences Center, Pittsburgh, PA for Gulf Oil Chemicals Co., Houston, TX
Other Last changed	Rev. 7/2/2001 (Prepared by a contractor to the Olefins Panel)